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An expedient synthesis of spiroketals: model studies for the calyculin C₁₆–C₂₅ fragment

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Abstract—A new short strategy to prepare the spiroketal fragment of calyculins is presented. A novel Seyferth–Gilbert type homologation of hindered lactols to the corresponding alkynes has been achieved for the first time. The spirocyclization was achieved efficiently via a DIHMA (double intramolecular hetero-Michael addition) process of this hindered ynone. The spirocyclization rate is not dependent on the stereochemistry of the alkoxy substituent in the oxolane ring.

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1. Introduction

The 1,6-dioxaspiro[4.5]decane ring system is a common motif, occurring in nearly 100 natural products. It is noteworthy that in most of these structures, the configuration of the stereogenic carbon atom is dictated by double anomeric effect, placing the oxygen in the oxolane ring axial with respect to the oxane ring (Fig. 1). Due to the wide occurrence of such structures, a rapid and reliable entry into the spirocyclic structure is highly desirable. This was of special interest to us because of our ongoing efforts towards the total synthesis of calyculin C, a potent protein phosphatase inhibitor. In this paper we report our recent results on a highly convergent strategy to achieve this goal.

Our retrosynthetic strategy for the model spiroketal is based on a convergent strategy (Scheme 1). The actual spiroketal formation is based on the DIHMA (double intramolecular hetero-Michael addition) process of a suitably derived ynone. Thus, our penultimate goal became the ynone 13a,b which would be available through a nucleophilic addition of the alkyne 8 onto the Weinreb amide 12, in turn available via Evans aldol methodology from propionyloxazolidinone 9 and benzyloxypropanal. The alkyne was envisioned to arise through a Seyferth–Gilbert-type homologation of the aldehyde (or lactol) corresponding to lactone 4.

Although seemingly well precedented, several questions remained to be answered. First, the electrophilic end of the ynone 13a,b is highly sterically crowded, which might affect the cyclization rate. Secondly, the formation of the highly substituted alkyne 8 is not trivial. Thirdly, the existence of the requisite alkoxy group in the oxolane ring might affect the cyclization rate and/or the stability of the ensuing spirocycle. To shed light on this latter question, we decided to enter the spirocyclization with enantiopure 12 and racemic 8. Rate differences between the diastereomers would thus become evident experimentally.

Keywords: Enantioselectivity; Natural product; Spiroketales.

Figure 1. Spiroketal fragment of calyculin C.
2. Results

The alkyne 8 was prepared as shown in Scheme 2, beginning with an addition of the ester enolate of ethyl isobutyrate to 2-benzoyloxyacetaldehyde 2 affording the hydroxy ester 3 in 63% yield. Protection of the hydroxy group (NaH, BnCl, 75%) and DIBAL-H reduction of the lactone gave lactol 5 in near quantitative yield, ready for the Seyferth–Gilbert type homologation to the alkyne without further purification. If the initial ester aldol reaction was allowed to warm to higher temperatures, the intermediate alkoide corresponding to 3 further reacted, by intramolecular benzoate transfer and ring closure, to give the hydroxy lactone directly. Quenching the reaction mixture with benzyl chloride gave 4 in a one-pot operation, however, with yields typically below 25%. We therefore decided to rely on the more reproducible two step operation.

Ohira’s reagent 6 is a mild alternative to the original Seyferth–Gilbert homologation, widely used to transform an aldehyde to the corresponding alkyne.8 In our case, the lactol 5 was used as the aldehyde surrogate.9 The relative sluggishness of the lactol for ring-chain tautomerism was evident experimentally: Ohira’s reagent 6 had to be added slowly (in five ca. 50 mol% portions over 5 days), and the reaction temperature had to be kept low (between 36 and 44 °C) in order to achieve acceptable yields reproducibly (60–79%, based on recovered starting material). Higher reaction temperatures or faster addition of reagent 6 and the base led to decomposed products. This successful procedure represents the first successful example of using a hindered lactol in the Seyferth–Gilbert homologation. Finally, the secondary hydroxyl was protected (TBSOTf, lutidine, 88%) to give the alkyne 8 ready for coupling.

The enantiopure fragment, Weinreb amide 12, was prepared using the diastereoselective Evans syn-aldol reaction from the known propionyloxazolidinone 9 and 3-benzoxypyrrolaldehyde (Scheme 3). Thus, reaction of the dibutylboron Z-enolate of 9 and the aldehyde gave the desired 10 in 95% yield. Conversion of 10 to the Weinreb amide 11 (82%), followed by TBS protection under standard conditions10 gave the coupling partner 12 (68%).

Fragments 8 and 12 were coupled using the Weinreb–Nahm procedure to produce alkynone 13a, b.11 Spirocyclization with the DIHMA procedure was then attempted using a stepwise protocol.11 In the first step, the TBS protections were cleavage by CSA in MeOH. Some spirocyclization occurred already at this stage (TLC). Thus, the solvent was removed and replaced with benzene, and addition of p-TsOH took the spirocyclization to completion. Because
the alkyne 8 was not optically pure, the two diastereoisomers 1a and 1b were observed in a 1:1 diastereomeric ratio (Scheme 4). This supports the conclusion that the cyclization rate is not critically dependent on the existence of a directing alkoxy group in the oxolane ring.

3. Conclusions

We have presented a new strategy to prepare the spiroketal fragment of calyculins. A novel Seyferth–Gilbert type homologation of hindered lactols to the corresponding alkynes has been achieved for the first time. The spirocyclization was achieved efficiently via a DIHMA (double intramolecular hetero-Michael addition) process. The spirocyclization rate is not dependent on the stereochemistry of the alkoxy substituents in the oxolane ring. Application of this protocol in the total synthesis of calyculins C will be reported in due course.

4. Experimental

4.1. General

All reactions were conducted under a positive pressure of argon. THF was distilled prior to use from sodium-benzophenone, MeOH from Mg(OMe)2 and toluene from argon. THF was distilled prior to use from sodium.

Melting points were determined on a Gallenkamp melting point apparatus MFB-595 and are uncorrected. TLC was conducted on Merck 0.25 mm silica gel 60 F plates and was visualized with UV light, anisaldehyde, PMA or ninhydrin staining. Flash chromatography was performed using Merck silica gel 60 (230–400 mesh) as a stationary phase. HPLC was performed with Waters 501 pump, Waters 486 tunable absorbance detector, Waters 746 data module and Resolve™ silica inserts for normal phase chromatography and Daicel Chiralcel OD 25 cm × 0.46 cm with Daicel Chiralcel OD 5 cm × 0.46 cm precolumn for chiral chromatography. Optical rotations were measured at 20 °C on a Perkin–Elmer polarimeter 343. IR spectra were measured with Perkin–Elmer Spectrum One.

Elemental analyses were performed with Perkin–Elmer Optima 4300 DV elemental Analyzer 2400 CHN. HRMS spectra were measured with Jeol JMS-DX 303 and Micromass LCT. NMR spectra were measured with Bruker AMX 400 (1H 400.13 MHz, 13C 100.61 MHz).

4.1.1. 3-(Ethoxycarbonyl)-2-hydroxy-3-methylbutyl benzolate 3. Diisopropylamine (2.82 mL, 20.1 mmol, 110 mol%) was dissolved in freshly distilled THF (20 mL) at 0 °C. BuLi (2.3 M, 8.7 mL, 20.1 mmol, 110 mol%) was added during 10 min and the light yellow solution was cooled to −78 °C. Ethyl isobutyrate (2.69 mL, 20.1 mmol, 110 mol%) was added dropwise during 5 min. The light yellow reaction mixture was stirred 1.5 h at −78 °C and aldehyde 2 (3.0 g, 18.3 mmol, 100 mol%) in THF was added dropwise over 20 min. After 2 h stirring at −78 °C, the reaction was quenched with sat. NH4Cl (20 mL) and allowed to warm up to rt. The aqueous phase was washed three times with 30 mL of Et2O, the combined organic phases were washed once with brine (20 mL) and dried with Na2SO4. The product was purified by flash column chromatography (30% EtOAc/hexane) affording 3 3.25 g (63%). Rf (50% EtOAc/hexane, UV/PMA) = 0.49; IR (νmax, film) 1141, 1366, 1386, 1581, 1598, 1737, 3565 cm−1; 1H NMR (400 MHz, CDCl3) δ 1.25 (t, J = 7.1 Hz, 3H), 1.29 (s, 3H), 1.32 (s, 3H), 3.15 (d, J = 6.6 Hz, 1H), 4.05 (m, 1H), 4.14 (dd, J = 7.1, 4.9 Hz, 2H), 4.37 (dd, J = 7.3, 11.7 Hz, 1H), 4.48 (dd, J = 2.9, 11.7 Hz, 1H), 7.74 (m, 2H), 7.57 (m, 1H), 8.06 (m, 2H); 13C NMR (100 MHz, CDCl3) δ 14.0, 20.9, 22.6, 45.4, 61.0, 66.2, 75.1, 128.4, 128.9, 129.7, 133.1, 166.7, 176.8; HRMS (TOF MS) calc for C15H21O5Na 303.1208, found 303.1221.

4.1.2. 4-(Benzyloxy)-dihydro-3,3-dimethylfur-2(3H)-one 4. NaH (60% oil dispersion, 476 mg, 11.9 mmol, 110 mol%) in dry DMF was cooled to 0 °C. Ester 3 (3.03 g, 10.8 mmol, 100 mol%) in THF (6 mL) was added dropwise at 0 °C. The reaction mixture was stirred for 5 min at 0 °C and 15 min at rt. BnCl (1.37 mL, 11.9 mmol, 110 mol%) was added dropwise and the reaction was stirred for 4 h at rt. After quenching at 0 °C with sat. NH4Cl, the aqueous phase was extracted three times with 25 mL of Et2O, the combined organic phases were washed once with brine (50 mL) and dried with MgSO4. After flash column chromatography (15% EtOAc/hexane) lactone 4 was isolated (1.77 g, 75%). Rf (30% EtOAc/hexane, UV/PMA) = 0.27; IR (νmax, film) 1773 cm−1; 1H NMR (400 MHz, CDCl3) δ 1.26 (s, 3H), 1.28 (s, 3H), 3.91 (dd, J = 4.0, 5.1 Hz, 1H), 4.15 (dd, J = 4.0, 10.1 Hz, 1H), 4.31 (dd, J = 5.1, 10.1 Hz, 1H), 4.59 (d, JAB =
11.1 Hz, 2H), 7.30–7.39 (m, 5H); 13C NMR (100 MHz, CDCl3) δ 17.9, 23.3, 42.9, 68.9, 72.1, 81.8, 127.5, 128.0, 128.5, 137.4, 180.6; HRMS (EI+) calcd for C13H16O3 220.1099, found 220.1092.

4.1.3. 4-(Benzyloxy)-tetrahydro-3,3-dimethylfuran-2-ol

Lactone 4 (0.273 g, 1.24 mmol, 100 mol%) in toluene (2.11 mL, 2.11 mmol, 170 mol%) was added during 5 min. After 14 min, the reaction was quenched with adding MeOH (0.5 mL) and allowed to warm up to rt. The solution was partitioned between 20 mL of 1 M HCl and 20 mL of EtOAc, the phases were separated and the aqueous phase was extracted three times with 15 mL of EtOAc. The combined organic phases were washed once with 10 mL of brine, dried with MgSO4 and evaporated affording crude 5 0.267 g, which was used without purification in the next reaction. Rf (50% EtOAc/hexane, UV/PMA) = 0.36; 1H NMR (400 MHz, CDCl3) δ 1.01 (s, 3H), 1.24 (s, 3H), 3.58 (d, J = 12.1 Hz, 1H), 3.61 (d, J = 3.7 Hz, 1H), 4.05 (dd, J = 10.5, 3.8 Hz, 1H), 4.23 (d, J = 10.2 Hz, 1H), 5.45 (d, JAB = 11.8 Hz, 2H), 4.81 (d, J = 11.9 Hz, 1H), 7.29–7.38 (m, 5H); 13C NMR (100 MHz, CDCl3) δ 17.4, 24.6, 46.6, 70.9, 72.0, 85.3, 105.3, 127.6, 127.9, 128.5, 137.4; HRMS (TOF MS EI+) calced for C8H8O3Na 241.1154, found 241.1180.

4.1.4. 2-Benzylx-3,3-dimethylpent-4-yn-1-ol 7.

A 25 mL 2-neck flask was charged with (12 mL) was cooled to 0 °C and di-tert-butylzinc reagent (1.1 mmol, 200 mol%) was added. After 12 min the reaction mixture was allowed to stir for 36 min before TBSOTf (0.253 mL, 1.1 mmol, 200 mol%) was added. After 12 min the reaction was quenched with 3 mL of sat. K2CO3. The mixture was warmed to 36 °C and allowed to stir for 5 days, during which more phosphonate 6 was isolated. The reaction mixture was cooled to −77 °C and 3-benzyloxy-propionaldehyde (6.2 g, 37.8 mmol, 110 mol%) dissolved in 10 mL CH2Cl2 was added slowly (45 min) keeping the internal temperature stable. Stirring was continued for a further 3 h at −77 °C and then for 30 min at 0 °C. Phosphate buffer (80 mL, pH 7.0) and methanol (60 mL) were added, and the mixture was cooled to −10 °C before slow (15 min) addition of 120 mL of (1:1) H2O2 (30%) and MeOH. The mixture was then stirred for 30 min at 0 °C before organic solvents were evaporated, Et3O was added and reaction was cooled to 0 °C. Sat. Na2S2O3 (120 mL) was added slowly (30 min) and the phases were separated. The aqueous phase was extracted three times with 80 mL Et3O and the combined organic phases were washed once with 80 mL of sat. NaHCO3 and 50 mL of brine and dried with Na2SO4. The crude product was purified by flash column chromatography (25% EtOAc/Hex) affording pure 10 (9.7 g, 71%). Rf (50% EtOAc/hexane, UV/PMA) = 0.31; [α]D20 = +44.7 (c 1.0; CHCl3); IR (νmax, film) 1111, 1694, 1780, 3480 cm−1; 1H NMR (400 MHz, CDCl3) δ 1.28 (d, J = 7.0 Hz, 3H), 1.74 (m, 1H), 1.89 (m, 1H), 2.78 (dd, J = 13.2, 9.5 Hz, 1H), 3.26 (dd, J = 13.5, 3.3 Hz, 1H), 3.29 (d, J = 2.6 Hz, 1H), 3.69 (m, 2H), 3.82 (dq, J = 7.0, 3.7 Hz, 1H), 4.18 (m, 1H), 4.19 (m, 1H), 4.52 (s, 2H), 4.68 (m, 1H), 7.34–7.37 (m, 10H); 13C NMR (100 MHz, CDCl3) δ 11.1, 33.7, 37.8, 42.6, 55.2, 66.1, 68.4, 70.4, 73.3, 127.4, 127.7, 128.4, 129.0, 129.4, 135.1, 138.0, 153.1, 176.7; HRMS (TOF MS EI+) calcd for C23H27NO5 397.1889, found 397.1880.

4.1.5. (2-(Benzyloxy)-3,3-dimethylpentanamide 11.

A 25 mL 2-neck flask was charged with N,O-Dimethyl hydroxylamine hydrochloride (1.08 g, 11.1 mmol, 220 mol%) and 4 mL THF. The suspension was cooled to −10 °C in a NaCl/ice-bath and AlMMe3 (5.3 mL, 10.6 mmol, 210 mol%) was added over 5 min. After 12 min, the cooling bath was removed and the reaction mixture was allowed to stir for 1 h at rt before it was cooled again to −10 °C. Oxazolidinone 10 (2.0 g, 5.0 mmol, 100 mol%) dissolved in a mixture (4:5) of CH2Cl2 (2.9 mL) and THF (3.5 mL) was slowly added.
The mixture was stirred for 1 h at 0 °C and then poured into a pre-cooled 0 °C mixture of aqueous HCl (0.5 M) (16 mL) and CH₂Cl₂ (16 mL). This was stirred for 1 h 15 min at 0 °C and the phases were separated. The aqueous phase was extracted three times with 60 mL of CH₂Cl₂ and the combined organic phases were washed once with 50 mL of brine and dried with MgSO₄. The crude product was purified by step gradient column chromatography (1:3, 2:5, 1:1 and 3:5 EtOAc/hexane/hexane in 900 mL fractions) affording 11 as a light yellow oil (1.16 g, 82%). Rₜ (50% EtOAc/Hex, UV/PMA) = 0.12; [α]D²θ = +11.4 (c 1.0; CHCl₃); IR (νmax, film) 1102, 1637, 3468 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 1.20 (d, J = 7.3 Hz, 3H), 1.89–1.66 (m, 2H), 2.93 (br s, 1H), 3.13 (s, 3H), 3.59 (dd, J = 5.4, 14.5 Hz, 2H), 6.4, 3.1 Hz, 1H), 3.67 (dd, J = 10.8, 7.1 Hz, 1H), 4.00 (d, J = 8.7, 7.8 Hz, 1H), 1.42 (dd, J = 7.7, 6.7 Hz, 1H), 2.43 (td, J = 9.2, 10.8 Hz, 2H), 2.78–2.75 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ 45.4, 4.4, 13.9, 6.9 Hz, 3H), 1.23 (s, 3H), 1.22 (s, 3H), 1.76–1.92 (m, J = 14.0, 6.4 Hz, 2H), 2.53–2.60 (m, J = 13.9, 6.9 Hz, 1H), 3.38 (dd, J = 7.1, 3.0 Hz, 1H), 2.59, 3.26, 5.8, 53.5, 53.5, 65.1, 65.1, 66.7, 70.0, 73.0, 74.5, 81.3, 85.5, 85.5, 99.3, 127.4, 127.4, 127.5, 127.7, 128.2, 128.3, 138.4, 138.7, 138.8, 190.2; HRMS (TOF MS El⁺) calcd for C₂₃H₂₅NO₃Si₂ 689.4034, found 689.4025.

4.1.10. (7R,8S)-3-Benzoxylol-7-(2-benzyloxy-ethyl)-4,4,8-trimethyl-1,6-dioxo-spiro[4,5]decan-9-one 1a and 1b. The mixture of ynones 13a.b (13 mg, 19.5 µmol, 100 mol%) was dissolved in 0.5 mL of dry MeOH and camphor sulphonic acid (0.7 mg, 3.0 µmol, 15 mol%) was added. The reaction was allowed to stir at rt for 2 h 20 min before the solvent was evaporated. The residue was dissolved in 1 mL benzene and the reaction was stirred for 3 h 30 min, after which time p-TsOH (1.4 mg, 7.4 µmol, 38 mol%) was added. Stirring was continued for another 15 h, and the reaction was quenched by adding TFA (0.02 mL) followed by 1 mL of sat. NaHCO₃. The phases were separated and the aqueous one was extracted three times with 3 mL of toluene. The combined organic phases were extracted once with 5 mL of brine and dried with MgSO₄. The crude product was first purified by step gradient column chromatography (5, 10, 15 and 20% EtOAc/hexane, 1:20, flow rate 1.0 mL/min, UV/PMA). The mixture of ynones 13a.b was dissolved in 0.5 mL of dry THF and cooled to 0 °C. DMAP (0.50 g, 4.19 mmol, 100 mol%) and imidazole (0.61 g, 7.3 mmol, 200 mol%) were added. After 50 min, the reaction mixture was allowed to stir at rt for 2 h 20 min. After another 2 h 15 min it was quenched with 5 mL of C₂H₅OH and the combined organic phases were washed once with 50 mL of brine and dried with MgSO₄. The crude product was purified by step gradient chromatography (150 mL 5% EtOAc/hexane, 150 mL 10% EtOAc/hexane/hexane) affording 13a.b (0.045 g, 79%). Rₜ (50% EtOAc/Hex, UV/PMA) = 0.66; IR (νmax, film) 836, 1103, 1663 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.01 (s, 3H), 0.04 (s, 3H), 0.06 (s, 3H), 0.07 (s, 3H), 0.84 (s, 9H), 0.91 (s, 9H), 1.13 (d, J = 6.9 Hz, 3H), 1.23 (s, 3H), 1.28 (s, 3H), 1.76–1.92 (m, J = 14.0, 6.4 Hz, 2H), 2.53–2.60 (m, J = 13.9, 6.9 Hz, 1H), 3.38 (dd, J = 7.1, 3.0 Hz, 1H), 2.59, 3.26, 5.8, 53.5, 53.5, 65.1, 65.1, 66.7, 70.0, 73.0, 74.5, 81.3, 85.5, 85.5, 99.3, 127.4, 127.4, 127.5, 127.7, 128.2, 128.3, 138.4, 138.7, 138.8, 190.2; HRMS (TOF MS El⁺) calcd for C₂₃H₂₅NO₃Si₂ 689.4034, found 689.4025.
(100 MHz, CDCl₃) δ 10.7, 16.6, 24.5, 29.7, 41.1, 49.0, 66.6, 67.0, 71.3, 72.5, 72.8, 85.4, 108.8, 127.1, 127.4, 127.5, 127.5, 128.3, 128.4, 138.6, 138.6, 210.4; HRMS (TOF MS EI⁺) calcd for C₂₂H₃₆O₂Na 461.2304, found 461.2317.

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References and notes


