Nimbalkar, Pranhita R.; Khedkar, Manisha A.; Chavan, Prakash V.; Bankar, Sandip B.

Enhanced biobutanol production in folic acid-induced medium by using clostridium acetobutylicum NRRL B-527

Published in:
ACS Omega

DOI:
10.1021/acsomega.9b00583

Published: 31/07/2019

Please cite the original version:
Enhanced Biobutanol Production in Folic Acid-Induced Medium by Using \textit{Clostridium acetobutylicum} NRRL B-527

Pranhita R. Nimbalkar,†‡ Manisha A. Khedkar,‡ Prakash V. Chavan,‡*† and Sandip B. Bankar‡*†

†Department of Bioproducts and Biosystems, School of Chemical Engineering, Aalto University, P.O. Box 16100, Aalto FI-00076, Finland
‡Department of Chemical Engineering, Bharati Vidyapeeth Deemed University College of Engineering, Pune 411043, India

\textbf{ABSTRACT}: The conventional acetone–butanol–ethanol fermentation process suffers from several key hurdles viz. low solvent titer, insufficient yield and productivity, and solvent intolerance which largely affect butanol commercialization. To counteract these issues, the effect of stimulator, namely, folic acid was investigated in the present study to improve butanol titer. Folic acid is involved in biosynthesis of a diverse range of cellular components, which subsequently alter the amino acid balance. Therefore, different concentrations of folic acid were screened, and 10 mg/L supplementation resulted in a maximum butanol production of 10.78 ± 0.09 g/L with total solvents of 18.91 ± 0.21 g/L. Folic acid addition at different time intervals was also optimized to get additional improvements in final butanol concentration. Overall, folic acid supplementation resulted in two-fold increase in butanol concentration and thus could be considered as a promising strategy to enhance solvent titer.

1. INTRODUCTION

Butanol has been extensively used as an industrial intermediate from past few decades and now attracted public attention for being an advanced biofuel.\textsuperscript{1} Based on fuel properties, butanol shows better comparability with gasoline as compared to different liquid fuels.\textsuperscript{2} However, the traditional acetone–butanol–ethanol (ABE) fermentation process has numerous pitfalls such as complex bioprocess, low butanol concentration, and end product inhibition.\textsuperscript{3} Hence, more research efforts in field of biobutanol production are needed to explore butanol as next-generation biofuel. Incidentally, various efforts including mutagenesis, metabolic engineering approaches, and integrated fermentation with competent product recovery were reported recently to enhance product titer.\textsuperscript{3} Moreover, the metabolic regulation to elevate butanol production has been carried out by incorporating organic acids and electron carrier together.\textsuperscript{3}

Several researchers have implemented certain precursors/cofactors/electron carriers and indeed, considerable successes have been achieved in raising butanol titer.\textsuperscript{5,6} Co-culturing of \textit{Clostridium acetobutylicum}/\textit{Saccharomyces cerevisiae} along with butyrate addition has maximized butanol concentration up to 16.3 g/L which is 46.8% higher than control.\textsuperscript{7} On the other hand, addition of calcium carbonate together with yeast extract promotes butanol synthesis to increase amino acid concentrations, and thereby stimulate cell growth.\textsuperscript{8} Similarly, butanol concentration can be upregulated by incorporation of substrate supplements such as butyrate, acetate, and allopurinol.\textsuperscript{9} Besides, the pH-controlled approach could be one of the promising techniques to increase sugar consumption rate and final solvent titer.\textsuperscript{10} Nonetheless, butanol toxicity during fermentation is again a key hurdle that significantly affects process yield and productivity. Therefore, augmenting strain robustness is of prime importance. Interestingly, upregulation of heat shock proteins could be helpful in adapting clostridial strains to high butanol concentration.\textsuperscript{11} Additionally, few amino acids that are either self-generated or exogenously incorporated, enhance the cell’s survival rate and butanol synthesis.\textsuperscript{12} Attempts were made in the current study to improve biobutanol production along with strain tolerance during batch fermentation by supplementing folic acid in production medium. Folic acid is mainly involved in the amino acid biosynthesis in microorganisms.\textsuperscript{13} The micro-organisms metabolically convert folic acid into tetrahydrofolic acid. This intermediate could then be diverted to amino acid or purine/thymidine synthesis. Hence, it was thought desirable to supplement folic acid in the fermentation medium, in order to induce amino acid generation during ABE biosynthesis. These self-generated amino acids in growing clostridia eventually trigger the biobutanol production. Furthermore, folic acid addition at different fermentation times was also investigated to improve the overall performance with respect to butanol/acetone (B/A) ratio.

Received: March 1, 2019
Accepted: July 18, 2019
Published: July 31, 2019

DOI: 10.1021/acsomega.9b00583
ACS Omega 2019, 4, 12978−12982
2. RESULTS AND DISCUSSION

2.1. Effect of Folic Acid on Biobutanol Production.
Folic acid is a stable synthetic form of folate, which is required by prokaryotic/eukaryotic cells for the production of cellular materials. Thus in present study, folic acid was incorporated in the clostridial ABE fermentation medium to investigate its stimulatory effect. It has been observed that precursors/cofactors/stimulators are very effective when present at low concentration in the fermentation medium. Also, our previous study demonstrated significant improvement in butanol titer when trace elements were supplemented in the concentration range of 1–100 mg/L. Therefore, folic acid was selectively incorporated in different concentrations as 1, 10, 50, and 100 mg/L. Figure 1 depicts ABE production under varied folic acid concentrations.

Interestingly, folic acid supplementation has positively triggered ABE fermentation, thus resulting considerable increment in total solvent production. Control (without folic acid) experiment produced around 6.48 ± 0.14 g/L of butanol with total ABE to be 11.44 ± 0.23 g/L. Conversely, 10 mg/L folic acid supplementation resulted in maximum butanol production up to 10.78 ± 0.09 g/L with ABE of 18.91 ± 0.21 g/L. Addition of folic acid beyond 10 mg/L led to drop in solvent titer, especially because of detrimental effects at higher concentration. Vink et al. also showed similar detrimental impact on Clostridium elegans ageing because of increased folate synthesis, and bacterial-dependent chronic toxicity.

The upsurge in solvent concentrations is may be due to stimulation of clostridial cells by folic acid for amino acid self-generation. It has been reported that effective amino acid assimilation yields higher butanol level along with a greater B/A ratio. The positive influence of folic acid was further evaluated by their addition at several fermentation time intervals.

2.2. Effect of Time-Wise Folic Acid Addition.
Optimized concentration of folic acid (10 mg/L) was incorporated at fermentation time intervals of 0, 4, 8, 24, and 31 h. The conventional clostridial growth profile shows that the exponential phase begins at 4–6 h which lasts until 32–36 h just before the stationary phase. C. acetobutylicum are known to enter in the solventogenic phase after 36 h to initiate ABE production. Therefore, the folic acid addition was not extended beyond 31 h. As can be seen in Figure 2, the highest butanol production (12.94 ± 0.09 g/L) was reached when folic acid was incorporated after 4 h of fermentation. However, further delay in addition time resulted into relatively lower solvent production. This is because when microorganisms just start to enter in the exponential growth phase, their cellular activities are at peak and additional folic acid can immediately be utilized in butyric acid production under a favorable acidogenic phase. Interestingly, notable changes were observed in acetone production, while ethanol production was slightly altered with different folic acid addition times. Acetone production was lowered when folic acid was added at 8 and 12 h of fermentation (Figure 2). The lowered acetone production was attributed to diversion of flux toward butanol.

Lowered acetone production obviously resulted in fluctuation in the B/A ratio in the presence of folic acid. From Figure 2, it was found that the maximum B/A ratio (2.27) was reached when folic acid was added after 8 h of fermentation. Although it was not evaluated in the current study, it was believed that the lowered acetone flux was eventually transferred to the butanol production pathway that ultimately improved the butanol titer. This is evident from the altered butanol/ethanol ratio, especially because of higher butanol production and steady ethanol accumulation. Overall, these findings clearly suggest that folic acid incorporation at the specific clostridial growth phase (beginning of exponential phase) is essential to elevate butanol titer. Besides, several researchers also pointed out the effectiveness of “precursor addition time” for increased biobutanol production. Furthermore, fermentation kinetics of control and folic acid-supplemented medium were also studied in order to assess the fermentation performance.

2.3. ABE Fermentation Profiles in the Presence of Folic Acid. Folic acid (10 mg/L) was added after 4 h of clostridial fermentation and ABE profile was studied with respect to growth, pH, and sugar consumption. The growth and production behavior of clostridia was significantly affected by addition folic acid with improved butanol yield and productivity of approximately two-fold. From Figure 3A, it was observed that clostridial growth in the presence of folic acid is consistently higher than control, throughout the fermentation. A lag period of ~4 h was observed, in which cells acclimatize to fermentation conditions. Thereafter, cells enter into exponen-
Folic acid supplementation resulted into rapid sugar consumption of around 19% higher than control (Figure 3A). This signification change the microbial phase behavior. Moreover, folic acid incorporation also aided increment in acetone and ethanol concentrations. Ding et al.20 reported that supplementation of folic acid also affected clostridia, it did not alter essential cellular components. Previously, folic acid addition was utilized as a stimulator to alter development and longevity via bacterial uptake of breakdown products.24 Besides, vitamins (A, B1, B2, B3, and C) are found to support bacterial growth which eventually improved butanol titer.26 Increasing these vitamin concentrations by 8-fold has triggered growth as well as the solvent formation rate. Similarly, other medium precursors have also been reported to elevate the butanol production positively. ABE fermentation was also carried out in a bioreactor supplemented with folic acid (after 4 h fermentation). As a result, butanol concentration of 12.31 ± 0.09 g/L and productivity of 0.14 g/L·h were achieved in batch fermentation with folic acid addition. The statistical analysis by using single-factor ANOVA was performed to evaluate significant difference between studied parameters. The p value < 0.05 demonstrated that the folic acid supplementation significantly affected clostridial growth, fermentation pH, and ABE production profiles positively. ABE fermentation was also carried out in a bioreactor supplemented with folic acid (after 4 h fermentation). As a result, butanol concentration of 12.31 ± 0.09 g/L with total solvents of 21.50 ± 0.20 g/L was obtained. Interestingly, folic acid traces were not detected after fermentation was over (120 h). This clearly implies that folic acid added during fermentation was utilized as a stimulator to alter essential cellular components. Previously, folic acid supplementation altered the C. elegans development and longevity via bacterial uptake of breakdown products.24 Besides, vitamins (A, B1, B2, B3, and C) are found to support bacterial growth which eventually improved butanol titer (21.56 g/L).25 Another study also revealed that two basic vitamins, namely, biotin and p-aminobenzoic acid are required to maintain active cell growth.26 Increasing these vitamin concentrations by 8-fold has triggered growth as well as the solvent formation rate. Similarly, other medium precursors have also been reported to elevate the butanol production (Table 1). This study will be continued further with several bioprocess modifications such as fed-batch and continuous process monitoring in order to see its large scale feasibility.
Table 1. Comparative Evaluation of the Present Study with Other Literature Reports

<table>
<thead>
<tr>
<th>precursors supplemented in ABE fermentation</th>
<th>butanol (g/L)</th>
<th>total ABE (g/L)</th>
<th>butanol yield (g/g)</th>
<th>references</th>
</tr>
</thead>
<tbody>
<tr>
<td>folic acid</td>
<td>12.94</td>
<td>23.23</td>
<td>0.24</td>
<td>current study</td>
</tr>
<tr>
<td>trace elements (nickel)</td>
<td>10.08</td>
<td>18.17</td>
<td>0.17</td>
<td>Nimbalkar et al.</td>
</tr>
<tr>
<td>butyric acid</td>
<td>18.05</td>
<td>20.17</td>
<td>0.50</td>
<td>Al-Shorgani et al.</td>
</tr>
<tr>
<td>yeast extract, NH4NO3</td>
<td>21.70</td>
<td>31.73</td>
<td>0.39</td>
<td>Abd-Alla and Elsadek El-Enany</td>
</tr>
<tr>
<td>cysteine, FeSO4, yeast extract</td>
<td>9.15</td>
<td>11.24</td>
<td>0.30</td>
<td>Diez-Antoliner et al.</td>
</tr>
<tr>
<td>methyl viologen, CaCO3</td>
<td>11.54</td>
<td>12.94</td>
<td>0.29</td>
<td>Kumar and Banerjee</td>
</tr>
</tbody>
</table>

3. CONCLUSIONS

The biobutanol process, although very old, is still struggling to be commercialized for its biofuel application, especially because of its lower production titers and end production inhibition. Hence, modification in metabolic stimulators like folic acid is especially important in order to divert the flux toward butanol production. Current work shows the importance of metabolic stimulators that affect the biobutanol production profile in clostridia with improved tolerance toward solvent toxicity. Butanol production stimulators such as folic acid increased the overall titer and fermentation performance. The folic acid supplementation yielded nearly two-fold rise in butanol concentration, thus reaching specific butanol yield of 0.24 g/g. This study also detailed an impact of folic acid addition time on butanol production and B/A ratio. Furthermore, fermentation profiles demonstrated that butanol production was greatly triggered in the stationary phase of clostridia. Maximum butanol concentration of 12.94 ± 0.09 g/L was achieved in presence of folic acid. Incidentally, the current study supported the butanol production pathway with addition of folic acid which is a precursor to many amino acid biosynthesis in microorganisms. The specific upsurge in butanol concentration with the help of folic acid would be an alternative and feasible approach which can be implemented with further modifications to strengthen ABE fermentation economics.

4. MATERIALS AND METHODS

4.1. Clostridial Strain and Its Maintenance. The bacterium, C. acetobutylicum NRRL B-527 was obtained from ARS (Agricultural Research Services) Culture Collection, USA. Freeze-dried cells were stimulated in sterile revival media (RM) containing (g/L): glucose (5.0), beef extract (10), yeast extract (3.0), peptone (10), sodium chloride (5.0), sodium acetate (3.0), soluble starch (1.0), L-cysteine hydrochloride (0.5), and resazurin (0.001), at pH 6.8. The RM culture bottles were incubated at 37 ± 2 °C for 48 h. Furthermore, the clostridial spore suspension was prepared according to Nimbalkar et al. and used whenever required.

4.2. Inoculum and Production Media. The seed culture was prepared by inoculating 2% (v/v) spores in sterile 80 mL-reinforced clostridial medium (RCM) through heat shock treatment. Further, the RCM culture bottles were kept in an incubator for 18–20 h at 37 °C. The fermentation medium (P2) used in the current study contained the following (g/L):

- glucose (60), ammonium acetate (2.2), dipotassium hydrogen phosphate (0.5), potassium dihydrogen phosphate (0.5), magnesium sulfate (0.2), p-aminobenzoic acid (0.1), thiamin (0.1), manganese sulfate (0.01), iron sulfate (0.01), sodium chloride (0.01), and biotin (0.01) at pH 6.5. The nitrogen was purged through fermentation media and autoclaved subsequently at 121 °C for 20 min. Analytical grade chemicals were used in the present study.

4.3. Fermentation Experiments. The batch experiments were carried out in 100 mL glass bottles with 80 mL production medium. Folic acid (at varied concentration range of 1–100 mg/L) was added in the production medium by filter sterilization before inoculation. Fermentation was initiated by inoculating 5% (v/v) actively growing clostridial cells and maintained at 37 °C for 120 h. The standard P2 medium was used as a control. Batch fermentation was also performed in a 3 L bioreactor (Dhruv Fabrotech, India) fitted with pH probe, temperature sensor, and working volume of 1.5 L. All other fermentation conditions for the bioreactor were kept similar as explained earlier unless otherwise stated. The effect of folic acid was determined by analyzing samples after each 24 h interval. The data points were plotted to compare the fermentation performance. All the experiments were performed at least in triplicate and results presented are with average value ±standard deviation (SD).

4.4. Analytical Techniques. Total solvents (acetone, butanol, and ethanol) and acids (acetic acid and butyric acid) were quantified by analyzing samples in gas chromatography (Agilent Technologies 7890B) equipped with AB-INNOWax capillary column (30 m × 0.32 mm × 1 mm) and flame ionization detector. The oven program was set as 90 (5 min hold) to 200 °C at 40 °C/min rise (5 min hold). The injector (0.5 μL injection volume) and detector were maintained at 200 and 250 °C, respectively. Unutilized glucose was determined by the dinitrosalicylic acid method. A UV–visible spectrophotometer (3000+, Labindia) was used to monitor clostridial growth by measuring optical density at 600 nm. Besides, a fermentation medium pH was also recorded with the help of a laboratory pH meter (Global, India). Folic acid concentration after fermentation was estimated according to Matias et al. Statistical analysis (ANOVA) was also performed to evaluate the significant difference between the parameters studied.

AUTHOR INFORMATION

Corresponding Authors
E-mail: pvchavan@bvucoop.edu.in. Phone: +91-020-24107390. Fax: +91-020-24372998 (P.V.C.).
E-mail: sandipbankar@gmail.com, sandip.bankan@aalto.fi.
Phone: +358 505777898. Fax: +358 9462373 (S.B.B.).

ORCID
Sandip B. Bankar: 0000-0003-0280-9949

Notes
The authors declare no competing financial interest.

ACKNOWLEDGMENTS

Authors are greatly thankful to the Department of Science and Technology (DST) of Ministry of Science and Technology, Government of India (DST INSPIRE faculty award, IFA 13-ENG-68/July 28, 2014) and IFRP-TEQIP-II (BVUCOE/2016-17/2808) for providing financial support during the course of this investigation.
REFERENCES


(7) Luo, H.; Zeng, Q.; Han, S.; Wang, Z.; Dong, Q.; Bi, Y.; Zhao, Y. High-efficient n-butanol production by co-culturing *Clostridium acetobutylicum* and *Sacccharomyces cerevisiae* integrated with butyrate fermentative supernatant addition. *World J. Microbiol. Biotechnol.* 2017, 33, 76.


