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ADAPTIVE

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ADAPTIVE: leArning DAta-dePendenT, concliSe molecular VEctors for fast, accurate metabolite identification from tandem mass spectra

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Abstract

Motivation: Metabolite identification is an important task in metabolomics to enhance the knowledge of biological systems. There have been a number of machine learning-based methods proposed for this task, which predict a chemical structure of a given spectrum through an intermediate (chemical structure) representation called molecular fingerprints. They usually have two steps: (i) predicting fingerprints from spectra; (ii) searching chemical compounds (in database) corresponding to the predicted fingerprints. Fingerprints are feature vectors, which are usually very large to cover all possible substructures and chemical properties, and therefore heavily redundant, in the sense of having many molecular (sub)structures irrelevant to the task, causing limited predictive performance and slow prediction.

Results: We propose ADAPTIVE, which has two parts: learning two mappings (i) from structures to molecular vectors and (ii) from spectra to molecular vectors. The first part learns molecular vectors for metabolites from given data, to be consistent with both spectra and chemical structures of metabolites. In more detail, molecular vectors are generated by a model, being parameterized by a message passing neural network, and parameters are estimated by maximizing the correlation between molecular vectors and the corresponding spectra in terms of Hilbert-Schmidt Independence Criterion. Molecular vectors generated by this model are compact and importantly adaptive (specific) to both given data and task of metabolite identification. The second part uses input output kernel regression (IOKR), the current cutting-edge method of metabolite identification. We empirically confirmed the effectiveness of ADAPTIVE by using a benchmark data, where ADAPTIVE outperformed the original IOKR in both predictive performance and computational efficiency.

Availability and implementation: The code will be accessed through http://www.bic.kyoto-u.ac.jp/pathway/tools/ADAPTIVE after the acceptance of this article.

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1 Introduction

Metabolites are small molecules, having many important functions in living cells such as energy transport, signaling, building blocks of cells and so on (Wishart, 2007). Identifying their biochemical characteristics or so-called metabolite identification is an essential task in metabolomics to increase the knowledge of biological systems. Yet, it is still a challenging task due to the size or coverage of spectra libraries.

Mass spectrometry (MS) is one of the most common techniques in analytical chemistry for dealing with metabolite identification (de Hoffmann and Stroobant, 2007). In more detail, a chemical compound is decomposed into fragments, of which mass-to-charge ratios (m/z) are continuously measured to obtain a mass spectrum. One MS spectrum can be represented by a list of peaks, each of which corresponds to a fragment captured by MS. Figure 1 shows a real example of a MS spectrum. In practice, tandem MS (also known as MS/MS or MS2) is widely used, in which precursor ions of specific m/z values from MS spectra are selected and further fragmented to produce other groups of product ions [see, e.g. Vaniya and Fiehn (2015) for more details]. The MS/MS spectra provide structural information about the measured compound, which makes MS/MS more useful for tackling metabolite identification.
A number of computational methods have been proposed for identifying unknown metabolites from MS/MS spectra data. In general, they are classified into three main categories: (i) spectral library search; (ii) in silico fragmentation; and (iii) machine learning (Nguyen et al., 2018a). Recent advances in metabolite identification have been led by the machine learning category (e.g. Brouard et al., 2016; Dührkop et al., 2015; Nguyen et al., 2018b). This category can be further divided into two key groups: supervised learning for substructure prediction and unsupervised learning for substructure annotation. While the former is to find a mapping from inputs (e.g. spectra) to outputs (e.g. fingerprints), the latter extracts underlying substructures of metabolites. Our research focuses on supervised learning, where the common scheme is to learn a mapping from spectra to structures.

The prediction can be divided into two steps: (i) fingerprint prediction: predicting fingerprints of a given test spectrum with supervised learning; (ii) candidate retrieval: retrieving chemical compound (from database) which is closest to the predicted fingerprints (Nguyen et al., 2018b).

Kernel methods have been shown to be effective for fingerprint prediction, such as methods include FingerID (Heinonen et al., 2012), CSI:FingerID (Dührkop et al., 2015) and input output kernel regression (IOKR, Brouard et al., 2016). In particular, IOKR is recognized as the current cutting-edge method for metabolite identification due to the following advantages: (i) structures (e.g. feature interaction in the molecular fingerprint vectors) in the output can be incorporated into the learning model by the kernel defined in the output space, leading to accuracy improvement; (ii) fingerprints are simultaneously predicted by the learned model, rather than being considered as a set of separate tasks, resulting in faster computation. One can take structures of the metabolites into account by using graph kernels (path, shortest-path and graphlet kernels) or kernels defined on molecular fingerprints. It is also known that kernels based on fingerprint vectors obtained the best performance (Brouard et al., 2016). However, a limitation of using molecular fingerprints as the intermediate representation vectors is that they are general-purpose and very large in size to encode all possible substructures and chemical properties related with metabolites. Consequently, such vectors are neither necessarily specific to any task nor data, and therefore redundant in the sense that these vectors might contain information irrelevant to the task, resulting in limited predictive performance. Moreover, the large size of fingerprints causes slow prediction in the first step of the above two steps.

Generally, in machine learning, deep learning has been proven successful recently in many application domains. Deep learning is useful for regular data, say a table, in which rows are instances and columns are features, and vice versa. However, semistructured data, particularly graphs, for example, chemical (or biological) molecules, which are irregular types of data, are difficult to be used with deep learning. A number of research efforts have been devoted to applying deep learning to semistructured data, proposing models to learn representations of graphs, such as Duvenaud et al. (2015), Li et al. (2015) and Nguyen et al. (2017). Importantly, Gilmer et al. (2017) showed that a lot of research on graphs can be formulated in a unified model, namely message passing neural network (MPNN), with the following three components: message passing, update and read-out functions. In other words, one way of defining such functions results in a different model for learning graphs. Furthermore, another attractive property of MPNN is that it allows to learn meaningful representations specific to each task for graphs in an end-to-end manner.

We propose a powerful machine learning framework for metabolite identification, named ADAPTIVE, which has two subtasks: (i) learning a mapping from structures to molecular vectors and (ii) learning a mapping from spectra to molecular vectors. Figure 2 shows a schematic picture of ADAPTIVE, where the left and right blue boxes correspond to the first and second subtasks, respectively. In Subtask 1, ADAPTIVE learns a model to generate molecular vectors for metabolites using their chemical structures, where these vectors are specific to both data and the task of metabolite identification, and therefore nonredundant. The model in Subtask 1 is parameterized by MPNN for mapping metabolite structures to the molecular vectors. The main contribution of this article is in the Subtask 1, that is, to learn the correspondence between given pairs of spectra and structures for metabolites.

Thus, the parameters of MPNN are trained so that the correlation between the spectra and the vectors mapped from the structures is maximized. We use Hilbert-Schmidt Independence Criterion (HSIC, Gretton et al., 2005) for evaluating the correlation, due to its theoretically nice properties and kernel-based calculation. Specifically, we formulate an objective function for the maximization problem through HSIC and solve this problem to have the best molecular vectors adapted to given data. For Subtask 2, ADAPTIVE uses IOKR to learn a mapping from spectra to molecular vectors generated by the Subtask 1.

We emphasize that the key difference between ADAPTIVE and the original IOKR is that IOKR uses ‘manually designed’ fingerprints, which are large in size, possibly redundant and nonspecific to metabolite identification (and given data), while ADAPTIVE learns representations for metabolites from given data, as molecular vectors, resulting in that the molecular vectors generated by ADAPTIVE are data-driven and concise.

In order to validate the performance of ADAPTIVE, we conducted extensive experiments using a benchmark data. Experimental results showed the following two main advantages of ADAPTIVE over existing methods, including the original IOKR:

- **Predictive performance**

ADAPTIVE achieved the best performance, followed by IOKR, CSI:FingerID and FingerID. For example, the top-20 accuracy of ADAPTIVE was 78.52% with the parameters of Gaussian kernel,
ALIGNF and molecular vector size of 300. On the other hand, IOKR, CSI:FingerID and FingerID achieved 74.79%, 73.07% (or 68.20%) and 58.17%, respectively, using Gaussian kernel (for IOKR) and ALIGNF. The top-$k$ accuracy was computed by the average over all trials of 10-fold cross-validation (CV), and so the performance advantage of ADAPTIVE was significant and very clear.

- Computational efficiency for prediction

Under the same experimental setting, ADAPTIVE was four to seven times faster than IOKR, which was already known as the fastest method. We can then say that ADAPTIVE is the current fastest method while keeping the highest predictive performance for metabolite identification.

2 Related work

As mentioned in the Introduction section, fingerprint prediction is important in supervised learning for metabolite identification, because we can retrieve metabolite candidates more reliably if fingerprints are predicted more accurately. For fingerprint prediction, kernel learning has been shown to be the most powerful approach. For example, a typical approach, FingerID (Heinonen et al., 2012) uses probability product kernel (PPK, Jebara et al., 2004), which can be directly computed from spectra and runs support vector machine with this kernel for solving fingerprint prediction as a classification problem. CSI:FingerID (Dührkop et al., 2015), an extension of FingerID, uses not only spectra but also fragmentation trees (FTs, Rasche et al., 2011) as input to generate kernels over spectra and FTs, which are then combined via multiple kernel learning (MKL, Gönen and Alpaydın, 2011). FTs may capture structural information behind spectra which is missing in the approach of FingerID. This is the motivation of CSI:FingerID. However, the computational cost for converting FTs from MS/MS spectra is very expensive, leading to heavy computational load, which causes a problem particularly in prediction. Thus, we can say that kernel-based supervised learning, particularly complex kernels, have a computation issue, regardless of high performance in prediction. On the other hand, a sparse learning model, namely SIMPLE (Nguyen et al., 2018b), considers a simpler function than kernels for fingerprint, while interactions of peaks in spectra can be incorporated into learning models explicitly. SIMPLE achieved a comparable performance against kernel-based learning, reducing the computational cost drastically. A key point of SIMPLE is to take advantage of sparsity of spectra, which results in faster prediction and interpretability, showing clear advantages over kernel-based methods.

Among the series of kernel-based approaches, IOKR (Brouard et al., 2016) has been shown to outperform the previous methods, in terms of both predictive performance and computational speed. It learns a mapping from spectra, i.e. input $X$, to molecular fingerprints (or structures behind fingerprints), i.e. output $Y$. In order to do this mapping, IOKR defines kernels to encode similarities in the input space (e.g. spectra and/or FTs) and the output space (molecular fingerprints or structures). Then, the advantage of IOKR comes from the following two points: (i) unlike previous kernel-based methods, IOKR handles the structured output space by the kernel defined for the output, which improves the predictive performance; (ii) IOKR simultaneously predicts fingerprints rather than considering fingerprint prediction as a set of separate tasks, leading to an efficient computation in prediction. Some part (mapping from spectra to feature vectors) of IOKR is a part of ADAPTIVE, and so further technical details of the corresponding part of IOKR is described more in Section 3.

Conventionally, molecular fingerprints for fingerprint prediction have been manually designed feature vectors to encode a predefined set of substructures or chemical properties, which are possibly found in metabolites. However, recently, machine learning-based (or data-driven) algorithms for generating fingerprints have been proposed. A typical approach is neural fingerprint (NFP, Duvenaud et al., 2015), which takes graphs with arbitrary sizes and shapes as inputs. NFP uses the idea of graph convolution, an extension of convolution operation from multidimensional arrays, like images or texts, to
3 Materials and methods

3.1 ADAPTIVE: overview

We first introduce the framework of ADAPTIVE for metabolite identification. This is also the general framework of approaches using machine learning for metabolite identification. It has two subtasks. Subtask 1: learning a function which maps metabolites from their structures to molecular vectors and Subtask 2: learning a function which maps metabolites from spectra to the vectors generated in Subtask 1. Figure 2 shows an illustration of the entire framework of ADAPTIVE. In this figure, the left and right blue boxes correspond to Subtasks 1 and 2, respectively.

For Subtask 1, given pairs of metabolite structure-spectrum, we estimate parameters of a function which maps metabolites from their structures to molecular vectors by maximizing the correlation between the vectors mapped from the structures and also the corresponding spectra. In more detail, we model the mapping function by MPNN and evaluate the correlation between the vectors and spectra by using HSIC due to the computational simplicity and provably theoretical properties of HSIC. For Subtask 2, we simply borrow the corresponding part of IOKR to learn a function mapping metabolites from spectra to vectors generated by Subtask 1.

We explain these two subtasks in the following subsections, being followed by the subsection on kernels we used in ADAPTIVE.

3.2 Subtask 1: learning molecular vectors for metabolites via HSIC

For this subtask, we need to estimate a function to map metabolites from structures to molecular vectors, given spectrum-structure pairs. For this problem, we use MPNN as the mapping function, which can extract meaningful representation for graphs (molecules for our problem) by supervised learning from training data. That is, MPNN requires labeled training data, which are, however, unavailable for this subtask. Then we manage this problem by taking advantage of given spectrum-structure pairs. We estimate parameters of MPNN by using the idea of maximizing the correlation between the given spectra and vectors (mapped from structures). The correlation is evaluated by HSIC. We describe the detail of MPNN, HSIC and related optimization procedures in the following subsections.

3.2.1 Message passing neural network

MPNN is a framework, which takes graphs of arbitrary sizes and structures as inputs, to learn their representation vectors at different levels (i.e. nodes, subgraphs and the whole graph) in a supervised manner (Gilmer et al., 2017). A key advantage is that MPNN allows to learn features specific to the given task from the given data. Below, we explain the procedure of MPNN.

First let G be an undirected graph, and ν and vw be a node (atom in molecules) and an edge (bond in molecules), respectively. Each node ν is assigned with state vectors at different levels, where each level represents a substructure (or subgraph) rooted at the corresponding node, denoted by hνr, where r shows a level. We can compute state vector hνr as well as message mνr by using the following two functions: message passing (1) and update (2):
is represented by feature vector, with only information of the node itself. We note that by repeatedly applying message passing and update functions (from left to right), more neighboring information are incorporated. For example, the updated feature (second level) has information on nodes 3 and 5, and then third level has that on nodes 2 to 5. Finally, the whole graph is covered

Importantly, estimation of HSIC is based on kernel calculation, which can effectively deal with the uncertainty of peaks in spectra caused by measurement errors.

Formally, we are given dataset $D = (X, Y) = \{(x_i, y_i)\}_{i=1}^n$, where $x_i, y_i$ are spectrum and molecular structure, respectively, of the $i$th metabolite. First, for the spectra, i.e. $x_i$, we consider kernels which combine spectra with FTs, namely $k(x_i, x_j)$. We describe the detail of the kernels for spectra in Section 3.4. Then, given the kernel over $X$ is fixed, the goal is to learn the function $\phi: \mathcal{Y} \rightarrow \mathcal{F}_d$ from $D$ such that the correlation between the input and output is maximized. The $\phi(y)$ is the output of MPNN (or molecular vectors) which belongs to space $\mathcal{F}_d$. The linear kernel function induced by this space can be written as follows:

$$I(y_i, y_j) = \langle \phi(y_i), \phi(y_j) \rangle$$

(4)

To evaluate the correlation between spectra and molecular vectors (output of MPNN), we use an unbiased empirical estimate of HSIC (Gretton et al., 2005), which can be given as follows:

$$u\text{HSIC}(X, Y) = \frac{1}{n(n-3)} \text{trace}(K_a L_a) + \frac{1}{(n-1)(n-2)} \text{trace}(K_a L_a L_a^T L_a)$$

$$- \frac{2}{n-2} \text{trace}(K_a L_a L_a)$$

(5)

where $K_a = K_a - \text{diag}(K_a)$ denotes the kernel matrix for the set of $n$ spectra $X$ with diagonal elements set to zero; $L_a$ is a vector of 1's of $n$ dimensions. Likewise $L_a = L_a - \text{diag}(L_a)$, where $L_a$ is the kernel matrix of $n$ molecular vectors output by MPNN. By arranging terms in (5), we can rewrite (5) as the objective function to learn parameters as follows:

$$u\text{HSIC}(X, Y) = \text{trace}(S_a L_a)$$

(6)
Learning adaptive molecular vectors for metabolite identification

However, directly optimizing (6) is prohibitively expensive in computation, particularly for large-scale data, since the complexity reaches $O(n^2)$, both in space and time. In order to overcome this limitation, following Zhang et al. (2018), we disjointly divide samples $(X, Y)$ into $n/B$ blocks with the size of $B$, $\{((x_{ib}^b, y_{ib}^b))_{i=1}^{n/B}\}_{b=1}^B$ and then apply HSIC on each block independently. An empirical estimate of the unbiased block HSIC can be defined by:

$$ubHSIC(X, Y) = \frac{1}{T} \sum_{i=1}^{n/B} \frac{1}{n/B} \sum_{b=1}^{n/B} \text{trace}(S_b L_b),$$

where $S_b$ can be defined by a similar manner to (7), and $L_b$ is the kernel matrix for the $b$th block.

Furthermore, in order to avoid the effect by biased partition of the dataset, following Yamada et al. (2018), we repeat shuffling dataset $T$ times, compute $ubHSIC$ on each permutation and take the average over them. HSIC by this procedure is known as bagging block HSIC, which can be written as follows:

$$ubHSIC(X, Y) = \frac{1}{T} \sum_{i=1}^{n/B} \frac{1}{n/B} \sum_{b=1}^{n/B} \text{trace}(S_{ib} L_{ib}).$$

We use (9) as objective function $f$ to learn parameters.

### 3.2.3 Optimization algorithm

An advantage of objective function (9) is that we can use the gradient descent (minibatch gradient descent) for estimating parameters of MPNN. We here explain details on how to conduct the minibatch gradient descent procedure for the HSIC-based loss, which has three steps.

**Step 1: Feed forward and loss calculation.**

For samples of size $n$, at each iteration, we perform random permutation and then split all samples into batches, where the size of each batch is $B$. Batches are sequentially fed into MPNN. The output of MPNN for the $b$th batch at the $t$th iteration is denoted by $\phi(Y_{ib}) = (\phi(y_{1ib}^b), \phi(y_{2ib}^b), \ldots, \phi(y_{nib}^b))$. Then using these outputs, the objective function on the whole samples can be calculated as in (9).

**Step 2: Gradient calculation of the loss layer.**

As we can compute the loss directly with the output of MPNN (i.e. $\phi(Y_{ib})$), we need to compute the gradient of $f$ with respect to $\phi(Y_{ib})$. Suppose that the output of MPNN is already normalized, i.e. $\phi(y_i^y) \phi(y) = 1$ for all $y \in Y$, the gradient can be obtained by the following:

$$\frac{\partial f}{\partial \phi(Y_{ib})} = B / T \sum_{b=1}^B \text{trace}(S_{ib} \phi(Y_{ib})).$$

**Step 3: Gradient calculation of the MPN and weight update.**

Having calculated the gradient of $f$, i.e. (10), the next step is to compute the gradient of $\phi(Y_{ib})$ with respect to model parameters $\theta$, namely $\frac{\partial \phi(Y_{ib})}{\partial \theta}$, to update the whole parameters for each batch at each step.

**Algorithm 2** is a pseudocode of the entire algorithm of learning parameters of MPNN.

### 3.3 Subtask 2: learning a mapping from spectra to molecular vectors by IOKR

For Subtask 2, we use IOKR. That is, we learn a mapping from spectra to molecular vectors generated in Subtask 1 by using IOKR. Again, we explain two technical reasons why we use IOKR for this mapping below: (i) IOKR allows to incorporate the structures behind outputs, such as feature interactions in molecular vectors, into the learning model, by which the prediction accuracy can be improved. (ii) Furthermore, all features in molecular vectors are predicted simultaneously, which is not like separate tasks in prediction. This leads to faster computation.

We now present the technical detail of IOKR below, which has two consecutive steps.

#### 3.3.1 Step 1: Learning spectra vectors mapping

Once parameters, i.e. function $\phi$, are learned, we convert the structures of metabolites into their molecular vectors to obtain a new set of pairs, $\{(x_i, \phi(y_i))\}_{i=1}^n$. Now the goal is to find the optimal function $h : X \to F_d$ by minimizing the following objective function:

$$\hat{h} = \arg\min_{h \in \mathcal{H}} \sum_{i=1}^n \|h(x_i) - \phi(y_i)\|^2_2 + \lambda \|h\|_{\mathcal{H}}^2,$$

where $\lambda (>0)$ is a regularization parameter to prevent overfitting and $\mathcal{H}$ is an approximate functional space that contains $h$; $F_d$ is a space of molecular vectors of dimension $d$.

By using the replresenter theorem in Micchelli and Pontil (2005), optimal solution $\hat{h}$ of (11) can be represented by a linear combination of vector-valued kernels on training set $X$:

$$h(x) = \sum_{i=1}^n \kappa\phi(x, x_i)c_i,$$

where $c_i (i = 1, \ldots, n)$ are vectors in $F_d$, $\kappa$ is an operator-valued kernel, defined on spectra $X$, satisfying certain constraints (see Michelli and Pontil, 2005). As dimensionality $d$ of space $F_d$ is finite, the kernel is a matrix with the size of $d \times d$.

By replacing $\hat{h}(x_i)$ in (11) with (12), $c_i (i = 1, \ldots, n)$ can be estimated in the following:

$$\text{vec}(C_n) = (\Lambda_{nd} + \kappa_n)^{-1}\text{vec}(\phi(Y_n)),$$

where $C_n = (c_1, c_2, \ldots, c_n)$ and $\phi(Y_n) = (\phi(y_1), \phi(y_2), \ldots, \phi(y_n))$ are both matrices with the size of $d \times n$, and $\text{vec}(\cdot)$ is the
vectorization of the input matrix, where the output is a vector obtained by repeatedly stacking each column of the input matrix on the top of the next column.

3.3.2 Step 2: Candidate retrieval
Given mapping $f$ learned in Step 1, we now turn to the problem of finding the output metabolite in the database which corresponds to the query spectrum $x$. To this end, we search metabolite $y$ in the list of given candidates $Y$, such that the squared distance between $\phi(y)$ and $b(x)$ can be minimized:

$$f(x) = \arg\min_{y \in Y} \| b(x) - \phi(y) \|^2_y,$$  \hfill (14)

Considering that the output kernel is normalized and the operator-valued kernel keeps $K_o(x, x') = k(x, x') \cdot I_t$, the optimal solution of $f(x)$ can be estimated as the following:

$$\hat{f}(x) = \arg\max_{\gamma \in Y} l(Y, \gamma)^T (I_{\gamma} + K_o)^{-1} k(X, x),$$  \hfill (15)

where $l(Y, \gamma) = \begin{bmatrix} l(y_1, \gamma) & \ldots & l(y_n, \gamma) \end{bmatrix}$ and $k(X, x) = \begin{bmatrix} k(x_1, x) & \ldots & k(x_n, x) \end{bmatrix}$ are column vectors.

Practically, the values given by objective function (15) are used as scores for ranking candidate metabolites in Step 2: candidate retrieval.

3.4 Kernels
ADAPTIVE uses kernels for the input and output.

3.4.1 Kernels for input
A various types of kernels are already defined and used for the input from MS/MS spectra. These kernels are typically divided into the following two groups: (i) kernels defined for spectra such as PPK (Jebara et al., 2004) and (ii) kernels defined for FPs (Rasche et al., 2011). Details on these kernels can be found in Dürrkop et al. (2015).

In fact, Dürrkop et al. (2015) suggested 24 different input kernels. ADAPTIVE combines these input kernels into a single kernel through MKL. (Gonen and Alpaydin, 2011). ADAPTIVE uses two options for MKL: (i) UNIMKL (uniform MKL): assigns the same weights to all component kernels, and (ii) ALIGNF: uses weights over kernels to combine. That is, in ALIGNF, weights over component kernels are optimized (trained) by maximizing the centered kernel alignment between the combined kernel and the target kernel defined on the molecular vectors, which generate trained parameters (model).

3.4.2 Kernels for output
After learning parameters (model) to generate the molecular vectors for structures, we define kernels for output $Y$ by directly computing kernels on the corresponding molecular vectors. In our experiments, we consider the following two typical kernels:

- Linear kernel: $l(y, y') = \phi(y)^T \phi(y')$.
- Gaussian kernel: $l(y, y') = \exp(-\gamma ||\phi(y) - \phi(y')||^2)$,

where $y$ and $y'$ are molecular structures in $Y$.

4 Experimental results
4.1 Dataset and evaluation measures
We used a benchmark dataset in Brouard et al. (2016) to evaluate ADAPTIVE and compare with existing methods. The dataset consists of 4138 MS/MS spectra extracted from the GNPS (Global Natural Products Social) public spectra library (https://gnps.ucsd.edu/ProteoSAFe/libraries.jsp).

To compare ADAPTIVE with existing methods, we used the same setting for all competing methods. Specifically we used 10-fold CV, and the results are averaged over all 10-folds. The performance was checked by the top-$k$ accuracies (where $k = 1, 10, 20$), which is the ratio of the number of the cases that the true structures are ranked at lower than or equal to $k$ to the number of all cases. Also the speed was checked by computation time for prediction, measured by milliseconds per example (ms/example).

Hyperparameters, such as regularization parameter $\lambda$ and parameter $\gamma$ of the output kernel, were chosen by using leave-one-out CV on each training fold. For prediction in ADAPTIVE, at the retrieval stage, given test example $x$, we computed the molecular vectors of $x$, $b(x)$ [see (11)] and those of all candidates $\phi(y)$ (see Algorithm 1). These candidates including the correct molecular structure of test example $x$ were ranked, according to their distances to $b(x)$ (from the smallest to the highest). These ranked candidates were used for computing the top-$k$ accuracy. Table 1 shows a set of parameter values, which were used to train MPNN of generating molecular vectors. State vectors of identical atoms at the lowest (atomic) level were initialized with the same random vector sampled from the standard normal distribution and updated during the training stage.

All experiments were performed on a server with 2.7 GHz Intel Core i5 CPU and 8GB memory. The code was written in Python and Matlab with the support of the Chainer framework (Tokui et al., 2015).

4.2 Performance results
4.2.1 Predictive performance
We compared the predictive performances of ADAPTIVE with three existing methods: FingerID (Heinonen et al., 2012), CSI:FingerID (Dürrkop et al., 2015) and IOKR (Brouard et al., 2016) in terms of the top-$k$ accuracy ($k = 1, 10$ and $20$). Table 2 shows the top-$k$ accuracies of the competing methods with UNIMKL and ALIGNF for MKL and linear and Gaussian kernels for the output kernel, changing $k$ from 1 to 20 and also changing the size of fingerprints from 100 to 300 (for ADAPTIVE only). This table first shows that ADAPTIVE achieved the best performance, being followed by IOKR, CSI:FingerID and FingerID. For example, ADAPTIVE with ALIGNF, Gaussian kernel and the fingerprint size of 300 achieved $31.03\%$ for $k = 1$, while IOKR with ALIGNF and Gaussian kernel was $29.59\%$ and CSI:FingerID with ALIGNF was $28.84\%$ or $24.82\%$. That of Finger: ID was only $17.74\%$. Interestingly, for $k = 1$, the performance advantage of ADAPTIVE against IOKR was rather slight, while $k = 10$ and $20$, ADAPTIVE outperformed IOKR much more clearly, with the difference of around 3–5% under the same condition for the two methods.
### Table 2. Comparison of the top-k accuracy (k = 1, 10 and 20) of FingerID, CSI:FingerID, IOKR and ADAPTIVE

<table>
<thead>
<tr>
<th>Method</th>
<th>Vec. size</th>
<th>MKL</th>
<th>Accuracies (mean/SD %)</th>
<th>Top 1</th>
<th>Top 10</th>
<th>Top 20</th>
</tr>
</thead>
<tbody>
<tr>
<td>FingerID</td>
<td>2765</td>
<td>None</td>
<td>17.74</td>
<td>49.59</td>
<td>58.17</td>
<td></td>
</tr>
<tr>
<td>CSI:FingerID</td>
<td>2765</td>
<td>ALIGNF</td>
<td>24.82</td>
<td>60.47</td>
<td>68.20</td>
<td></td>
</tr>
<tr>
<td>CSI:FingerID</td>
<td>2765</td>
<td>ALIGNF</td>
<td>28.34</td>
<td>66.07</td>
<td>73.07</td>
<td></td>
</tr>
</tbody>
</table>

Note: The highest value (indicating the most accurate prediction) are in boldface for each k.

### Table 3. Computation time for prediction by ADAPTIVE and IOKR

<table>
<thead>
<tr>
<th>Method</th>
<th>Mol. vec. size</th>
<th>Linear</th>
<th>Gaussian</th>
</tr>
</thead>
<tbody>
<tr>
<td>IOKR</td>
<td>2765</td>
<td>140.22</td>
<td>3352.4</td>
</tr>
<tr>
<td>ADAPTIVE</td>
<td>100</td>
<td>20.32</td>
<td>802.6</td>
</tr>
<tr>
<td></td>
<td>200</td>
<td>39.88</td>
<td>844.33</td>
</tr>
<tr>
<td></td>
<td>300</td>
<td>54.14</td>
<td>1071.8</td>
</tr>
</tbody>
</table>

Note: The smallest values (indicating the fastest) were in boldface for linear and Gaussian kernels.

### 4.3 Case study

To understand the results obtained by ADAPTIVE more, in the obtained molecular vectors, we examined substructures rooted at atoms, which activated several example features most. As shown in Section 3.2.1, each substructure rooted at an atom is represented by a state vector and contributes to computing the molecular vector of the whole molecule. Then, given a feature, we can estimate the contribution of each substructure by simply computing the softmax value from the corresponding state vector. We use these values of substructures as scores to rank substructures to activate the given feature.

Figure 5 shows three example features (#2, #39 and #83). For each feature, we show three substructures with the highest scores (each score is shown above the substructure). The first row shows three substructures which activated feature #2 most. Interestingly, we can see that these substructures share a smaller similar group of atoms: O, P and S (highlighted in blue). Similarly, the second row shows three substructures sharing a group of atoms: O and N, where these substructures activated feature #39 most. Also the third row shows substructures which activated feature #83, all having atom: Cl. Thus, Figure 5 shows that each feature of ADAPTIVE is activated by multiple different substructures sharing some similar properties, which must be important in data and probably for prediction. In contrast, each feature in regular molecular fingerprints is activated by only one predefined substructure. In summary, from this case study, learned features of ADAPTIVE are more concise and specific to the task of metabolite identification than regular molecular fingerprints, leading to the advantage of predictive performance and computation time.

### 5 Discussion and conclusion

Supervised learning for metabolite identification uses fingerprints as intermediate representation vectors between spectra and metabolites, while such fixed vectors are too redundant to cover all possible substructures and chemical properties in metabolites, causing limitations in predictive performance and high computational costs. To overcome this problem, we have proposed ADAPTIVE, which generates representations of metabolites specific to given spectrum-structure pairs. ADAPTIVE learns a model to generate molecular vectors for metabolites, which is parameterized by an MPNN over given molecular structures and trained through optimizing the objective function to maximize the correlation between molecular vectors and corresponding spectra. Our empirical validation of ADAPTIVE with the benchmark dataset showed the advantage of ADAPTIVE over existing methods including IOKR, the current cutting-edge method, both in predictive performance and computation time for prediction.

A drawback of ADAPTIVE would be interpretability, because structural information is implicitly encoded in compact vectors in ADAPTIVE and cannot be made explicit easily. In metabolite identification, it would be desirable to connect the set of peaks to the whole molecule. Then, given a feature, we can estimate the contribution of each substructure by simply computing the softmax value from the corresponding state vector. We use these values of substructures as scores to rank substructures to activate the given feature.
corresponding substructures/chemical properties of metabolites (Nguyen et al., 2018b). Developing a model with such interpretability would be interesting future work.

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References


Fig. 5. Example features (#2, #39 and #83) and their three substructures (and their scores) which activated the corresponding feature most. Note that three substructures of each feature share a similar group (set) of atoms which are shown in blue.