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Rapid Cationization of Gold Nanoparticles by Two-Step Phase Transfer**

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Abstract: Cationic gold nanoparticles offer intriguing opportunities as drug carriers and building blocks for self-assemblies. Despite major progress on gold nanoparticle research in general, synthesis of cationic gold particles larger than 5 nm remains a major challenge, though they would give tremendously enhanced plasmonic response. Here we present the first synthesis of cationic gold nanoparticles with tunable size between 8–20 nm prepared by a rapid two-step phase transfer protocol starting from simple citrate-capped particles. These cationic nanoparticles form ordered self-assembled structures with negatively charged biological components through electrostatic interactions.

Among metallic nanoparticles, gold nanoparticles (AuNPs) have been most extensively studied due to their high stability and intriguing optical properties, dominated by the localized surface plasmon resonance (LSPR).\cite{1−3} Because of their size, plasmonic properties and functionalization possibilities, they have potential in applications related to, for example, gene and drug delivery,\cite{4−7} biodiagnostics,\cite{7−9} detection of metal ions or biomolecules,\cite{6−10} and imaging.\cite{5−6} Importantly, the properties and possible applications of AuNPs strongly depend on their surface functionalization. Considering the vast amount of literature on AuNPs, there are relatively few reports on preparation of cationic AuNPs,\cite{11−17} even though they are interesting candidates especially for gene and drug delivery applications as well as for various self-assembled structures where uniform size of building blocks is essential.\cite{18−19} Thus, development of synthesis strategies aiming for cationic AuNPs with excellent size control, colloidal stability and narrow particle size distribution (PSD) is of high importance.

Previously, cationic AuNPs have been mainly prepared either by 1) directly reducing gold salts in presence of cationic ligands\cite{11−17} or 2) through place-exchange reactions or covalent linking of cationic ligands to AuNPs prepared by Brust-Schiffrin method.\cite{20−22} Out of these methods, the size control and resulting particle monodispersity of the first method are typically poor and the second method is limited to particle sizes of ≤ 5 nm leading to only weak LSPR (Figure 1). Larger nanoparticles would offer stronger light absorption in the visible range due to the approximately cubic dependence of the LSPR intensity with particle size.\cite{23} The strong and narrow LSPR absorption observed with large AuNPs would provide a more desirable starting point for e.g. AuNP-based sensing applications.

One of the simplest methods to produce anionic AuNPs larger than 5 nm with a narrow PSD is the citrate reduction method, pioneered in 1951.\cite{24} Citrate-AuNPs are straightforward to produce and their size can be tuned by varying the Au/citrate ratio,\cite{25} reaction solution pH or temperature,\cite{26} and via seeded growth approaches.\cite{27} Because of their fast, straightforward and well-studied synthesis and high nanoparticle yield, they are tempting precursors for further functionalization efforts. For these reasons, citrate-AuNPs have been functionalized with stabilizing ligands including various amines\cite{28} and thiols such as thiolated DNA,\cite{29} polymers containing thiol or disulfide groups,\cite{30,31} and small thiol molecules, e.g. mercaptosuccinic acid,\cite{32} cysteine,\cite{33} thiocysteine,\cite{34} or hydrophobic thiols.\cite{35} However, most of these systems produce negatively charged AuNPs because direct functionalization with a positively charged ligand typically leads to unwanted aggregation of AuNPs due to the detrimental electrostatic attraction between the ligands (Figure S1).

In this communication, we present the first synthesis of cationic gold nanoparticles with tunable size between 8–20 nm and narrow size distribution. The procedure involves a simple and rapid two-step phase transfer utilizing amine and thiol ligands. The resulting AuNPs are protected by thiols carrying quaternary ammonium groups making the AuNPs positively charged in a wide pH range. The two-step functionalization can be performed directly after the synthesis of citrate-AuNPs, is scalable, and the whole cationization process can be completed in less than an hour. To demonstrate their potential to efficiently bind large biomolecules and act as building blocks for nanoscale assemblies, we show that the cationic AuNPs readily interact with negatively charged virus particles and pack them into ordered assemblies.

Five different sizes of citrate-AuNPs (batches #1–#5) were synthesized according to a modified Turkevich method.

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Supporting information for this article is given via a link at the end of the document.

Figure 1. a) UV-vis spectra of different cationic AuNPs: 2.6 nm (red; made by Brust-Schiffrin method), 8.5 nm (green; made by two-step phase transfer) and polydisperse mixture (blue; made by direct synthesis). The molar nanoparticle concentration was adjusted to be the same in 2.6 and 8.5 nm AuNPs to illustrate the difference in their LSPR strength. b) Corresponding TEM images, scale bars equal 20 nm.
The citrate-AuNPs were cationized by a two-step phase transfer procedure shown schematically in Figure 2. Briefly, 30 mL of citrate-AuNPs were transferred to 6 mL of toluene by adding 5 µmol octadecylamine (ODA) and by shaking the two-phase mixture vigorously. The colored organic phase was separated and washed with water to remove possible remnants of citrate ions. Thereafter, 3 mL water and 300 µL (11-mercaptoundecyl)-N,N,N-trimethylammonium bromide (MUTAB, 4 mM in ethanol) were added to form a biphasic system and the tube was shaken to initiate the transfer of ODA-AuNPs to the aqueous phase. The transfer was completed by acidifying the mixture by HCl, which causes protonation of ODA and thus detaches the remaining ODA molecules from the AuNP surface. Depending on the extent of excess ligands and pH of the aqueous phase, ODA and MUTAB can act as surfactants, and occasionally an emulsion was formed. This gel-like material could be localized to the liquid-liquid interface by centrifugation and the aqueous phase could be easily collected. The aqueous phase was repeatedly washed with toluene to remove ODA and MUTAB residues. 1H-NMR spectroscopy was used to confirm the ligand compositions of amine- and thiol-protected AuNPs (Figure S2).

The amounts of ligands used in the procedure were optimized for the citrate-AuNP batch #1 (see Figure S3 and S4). We found that the optimal ODA amount was 0.8 times the amount of Au atoms used in the synthesis. By evaluating the size of the AuNPs with transmission electron microscopy (TEM) and estimating the number of their surface atoms (Supporting information),[23] this ODA amount corresponds to roughly 5 times the surface atoms of the AuNPs. On the other hand, the amount of MUTAB was optimized to 0.25 times the amount of Au atoms which corresponds to roughly 1.75 times the surface atoms of the AuNPs. Importantly, this phase transfer method allows increasing the concentration of AuNPs during the cationization process by simply adjusting the volumes of the phases. Over 30 times increase in concentration can be easily realized when starting with a large volume of citrate-AuNPs (Figure S5). The MUTAB-AuNPs can be concentrated even further by centrifugation, reaching concentrations over 20 mg mL⁻¹.

The cationization strategy is based on differences in Au-ligand interactions of citrate, amine and thiol ligands. Citrate ligands are only electrostatically bound to the AuNP surface while the strength of interaction increases with amine ligands and furthermore with thiol ligands, which form covalent Au-thiolate bonds. Importantly, a neutral amine (ODA) capping is utilized to avoid detrimental electrostatic interactions between the oppositely charged ligands. In addition to ODA, also the less expensive oleylamine can be utilized in the phase transfer.[36] It is worth noting that direct exchange of citrate to an alkanethiol in toluene causes an irreversible precipitation of AuNPs, thus making the “semistable” amine-capping indispensable (Figure S6). Finally, the thiolated MUTAB-AuNPs are highly stable and can be stored for months without notable aggregation. Analysis of the AuNP batch #1 UV-vis spectra (Figure 3) shows that the LSPR maximum shifts only very slightly from citrate-AuNPs to MUTAB-AuNPs suggesting that particle aggregation is negligible. Similar behavior is observed in all AuNP batches #1–#5 (Figure S7). The slight red shift of the LSPR in MUTAB-AuNPs is suggested to arise from the change in the dielectric constant of the ligand layer. The red shift is more prominent in ODA-AuNPs and it is enhanced by the higher refractive index of toluene compared to water.

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A further simplification to the procedure is that it is not critical to pinpoint the exact amounts of ligands needed for transfer. Incoming ligands can be gradually added and the completion of the transfer can be visually observed from the color of the phases (Figure 3B). The gradual addition of ligands only slightly decreases the final yield of the process (Figure S3B). It is also worth noting that the efficiency of the second phase transfer step is affected by the material of the reaction vessel and thus should be done in a plastic vessel to avoid MUTAB-AuNPs sticking to the negatively charged glass walls. This two-step procedure was found to be nearly quantitative for the smallest particles in this study (batch #1, 8.5 nm). For larger particles, the typical yield was approximately 50-80%, with most loss occurring during the first phase transfer step.

In addition to quaternized ammonium moieties, we have also prepared AuNPs functionalized with primary amines by using 6-aminohexanethiol hydrochloride (AHT) in the second exchange. In this case, the phase transfer process happened more slowly and needed several hours to complete. The UV-vis spectrum of the AHT-AuNPs was comparable to the spectrum of MUTAB-AuNPs (Figure S8).

In order to further verify that the phase transfer process does not change the size of the AuNPs, we imaged the particles before and after the cationization with TEM and determined the PSDs (Figure 4 and S9). As seen from the PSDs, there are no changes in the particle core sizes in batches #1–#3. The PSD in batch #4 gets slightly narrower in the cationization process. In order to probe the upper particle size limit of the procedure, batch #5 was deliberately synthesized to have a wider PSD ranging from 15 to 28 nm. As seen from the PSD data of batch #5, the efficiency of the phase transfer drops substantially above 20 nm. Based on the visual observations during the phase transfer experiments, most loss of particles occurs due to aggregation during the first transfer from citrate-AuNPs to ODA-AuNPs. Thus, citrate-AuNPs larger than 20 nm do not readily undergo the first phase transfer, likely due to the lower curvature of the larger particles leading to poorly protected and easily aggregating AuNP surfaces at the liquid-liquid interface.[28] Therefore, this procedure is not directly suitable for cationization of >20 nm nanoparticles.

In addition to TEM, dynamic light scattering (DLS) was also used to analyze the PSDs (Figure 3). For citrate-AuNPs, the average hydrodynamic diameters ($D_h$) were 2–3 nm larger than the average core diameter ($D_{core}$) measured by TEM. Similarly, the difference between $D_{core}$ and $D_h$ in MUTAB-AuNPs was 4–6 nm. These changes in $D_h$ reflect the sizes of the ligands and are thus consistent with the $D_{core}$ values obtained by TEM.

We also investigated the zeta potentials ($\zeta$) of AuNPs with DLS. As expected, the average zeta potentials of the citrate-AuNPs were negative and ranged from −42.8 to −72.6 mV, slightly depending on the particle size. In contrary, the average zeta potentials of MUTAB-AuNPs were positive and varied from +40.5 to +63.2 mV, indicating a successful cationization of AuNPs. The zeta potential distributions of AuNP batches #1–#5 are given in Table S1. To verify the application potential in biological systems, the colloidal stability of MUTAB-AuNPs was tested in 10 mM phosphate buffered saline (PBS). No change in the LSPR was observed during 20 hours incubation indicating high colloidal stability (Figure S11). In order to utilize this high stability and to
demonstrate the potential of these particles to bind biologically relevant macromolecules, we prepared bio-templated electrostatic self-assemblies of MUTAB-AuNPs by combining them with the cowpea chlorotic mottle virus (CCMV) particles. The oppositely charged particles rapidly self-assembled generating visible complexes in less than 10 minutes. The structure of the colloidal assemblies was further characterized using small angle X-ray scattering (Figure S12) indicating the formation of a AB3 face-centred-cubic (fcc) structure, in contrast with the AB8 crystal structure observed with smaller 2.6 nm cationic AuNPs in an earlier study. Here, the use of large (> 8 nm) cationic AuNPs allows the preparation of complexes with hybridized nanoparticle plasmon modes, which have previously been inaccessible with small AuNPs.

In conclusion, we have developed a facile, rapid and scalable cationization strategy for gold nanoparticles of sizes from 8 to 20 nm using simple citrate-AuNPs as precursors. Cationic nanoparticles in this size range are interesting candidates for applications related to plasmon enhancement and for building blocks in biohybrid assemblies.

Keywords: functionalization • gold • nanoparticles

Cationic gold nanoparticles offer intriguing opportunities as drug carriers and building blocks for self-assemblies. Despite major progress on gold nanoparticle research in general, synthesis of cationic gold particles larger than 5 nm remains a major challenge, though they would give tremendously enhanced plasmonic response. Here we present the first synthesis of cationic gold nanoparticles with tunable size between 8−20 nm prepared by a rapid two-step phase transfer protocol starting from simple citrate-capped particles. These cationic particles form ordered self-assembled structures with negatively charged biological components through electrostatic interactions.